



New records of Phallales (Basidiomycota) from Brazilian semi-arid region

Lima AA², Gurgel RAF¹, Oliveira RL², Ferreira RJ³, Barbosa MMB³ and Baseia IG^{1,2,3*}

¹ Departamento de Botânica e Zoologia, Centro de Biociências, Universidade Federal do Rio Grande do Norte, Av. Senador Salgado Filho, 3000, Campus Universitário, Natal, 59072-970, RN, Brazil

² Programa de Pós-Graduação em Sistemática e Evolução, Centro de Biociências, Universidade Federal do Rio Grande do Norte, Av. Senador Salgado Filho, 3000, Campus Universitário, Natal, 59072-970, RN, Brazil

³ Programa de Pós-Graduação de Biologia de Fungos, Centro de Ciências Biológicas, Universidade Federal de Pernambuco, Av. Prof. Moraes Rego, 1235, Cidade Universitária, Recife, PE, 50670-901, Brazil

Lima AA, Gurgel RAF, Oliveira RL, Ferreira RJ, Barbosa MMB, Baseia IG 2019 – New records of Phallales (Basidiomycota) from Brazilian semi-arid region. Current Research in Environmental & Applied Mycology (Journal of Fungal Biology) 9(1), 15–24, Doi 10.5943/cream/9/1/2

Abstract

Molecular studies have shown that Phallales constitutes a distinct clade within Phallomycetidae and morphologically comprises a wide variety of patterns of shapes and colors, producing expanded basidiomata, commonly known as stinkhorns, which may be free, latticed or sequestrate. Aiming to increase the knowledge of these fungi in the Brazilian semi-arid region, several fieldtrips were conducted in the rainy season of 2014 and 2018 in the Araripe National Forest, which covers an area of about 38,262 ha and is located in the State of Ceará. Several specimens were macro and micro morphologically described, and five species were identified. *Laternea dringii* is the second record outside of Mexico (locality type), *Mutinus albo truncatus* is the second record for science, *Mutinus argentinus* and *Phallus indusiatus* are the first records for the Brazilian semi-arid.

Key words – Biodiversity – cage-fungi – gasteromycetes – stinkhorns – taxonomy

Introduction

Phallales E. Fisch. is a molecularly well-defined order within Phallomycetidae Hosaka, Castellano & Spatafora (Hosaka et al. 2006, Hibbett et al. 2014) and morphologically comprises a wide variety of patterns of shapes and colors, producing expanded basidiomata, commonly known as stinkhorns, which may be free, latticed or sequestrate, such as *Claustula* K.M. Curtis, *Gelopellis* Zeller, *Protuberata* Möller and *Restingomyces* Sulzbacher, Grebenc & Baseia (Sulzbacher et al. 2016). Basidiospore dispersion generally occurs through a variety of mycophagous animals, especially flies that are attracted by the odor of the mature gleba (Tuno 1998).

According to Dring (1980), the tropics are considered the center of diversity for many of the species in this group. In particular, Brazil is represented by approximately 51 taxa of phalloid fungi. However, some of these records are considered synonyms or doubtful records because the voucher specimens are not well preserved (Cortez et al. 2011a, b).

Covering nine states, the Brazilian semi-arid occupies 982,563 km² of the national territory

(Pereira-Junior 2007), an area larger than countries such as Nigeria, Tanzania and Venezuela. In this area there is a predominance of the Caatinga, an exclusively Brazilian biome (Leal et al. 2005), as well as other vegetative enclaves that make the Brazilian semi-arid present a great diversity of plant species, many of them endemic to the biome. Through mycobiota studies in this region, it has been shown to be an interesting environment for the development of phalloid fungi, including the discovery of new taxa for science (Fazolino et al. 2010, Silva et al. 2015, Crous et al. 2017). The main goal of the present study is to contribute to an increase in the knowledge about gasteroid fungi in the Brazilian semi-arid region, based on morphological data.

Materials & Methods

Studied area

The Araripe National Forest covers an area of about 38,262 ha and has a perimeter of about 138 km, occupying part of the municipalities of Barbalha, Crato, Jardim and Santana do Cariri, all located in the State of Ceará (MMA 2006). The collections were carried out in the municipality of Crato (latitude 7°13'46" S and longitude 39°24'32" W), where habitat fragmentation is observed on roads and trails that cut through the preserved area (IBGE 2010). Fieldtrips occurred during the rainy season (February to May) of the years 2014 and 2018 (Fig. 1).

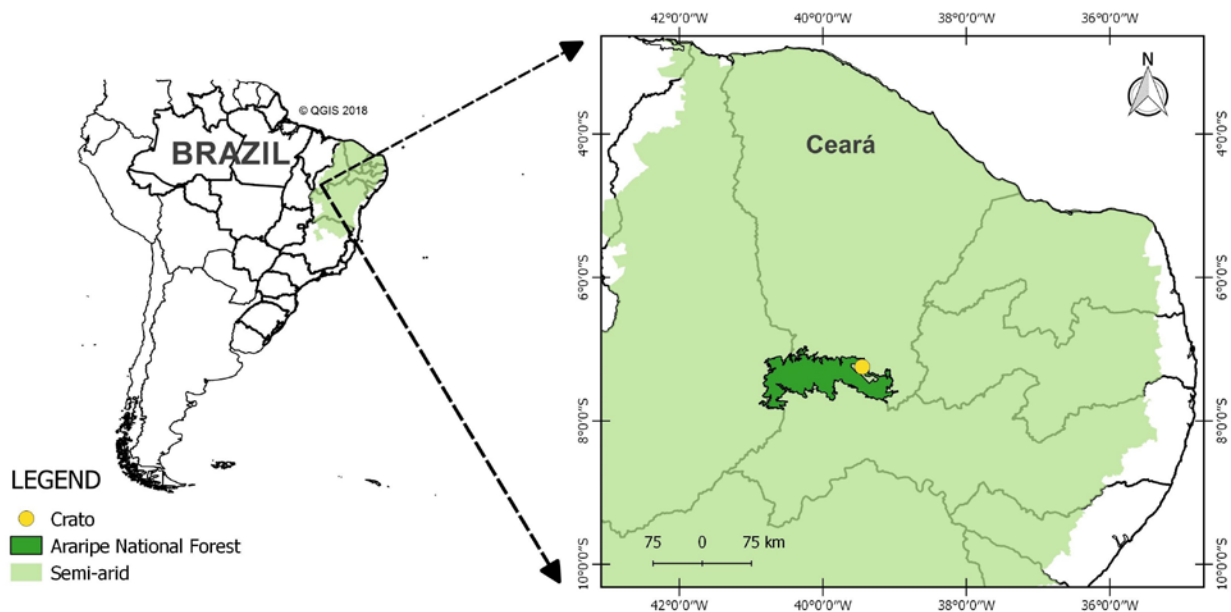


Fig. 1 – Map of the study area.

Collection methods

The specimens were collected following the methodology used by Baseia et al. (2014), in which the basidiomata were photographed with scales and geographical data and were removed from the substrate with a pocketknife. In the Biology of Fungi lab at the Federal University of Rio Grande do Norte the basidiomata were dried in an electric dehydrator and packaged with a ziplock bag; these packs were then placed together with an information sheet from the collection. The specimens were deposited in the Fungi Collection of the Federal University of Rio Grande do Norte (UFRN), Natal, Brazil.

Morphological observations

The morphological analysis was conducted according to Baseia et al. (2014). Macroscopic characteristics are based on observations of the fresh basidiomata, including characteristics of the

peridium and gleba. The size, shape and color of these characters were analyzed. Microscopic observation was performed using an optical Nikon Eclipse Ni (LM) microscope with Nikon DS-Ri1 camera attached, and all measurements were carried out using the NIS-Elements AR v.4.51.00 software. Slides for microscopic analyses were mounted in 5% KOH and Congo red containing portions of the fertile part, volva, receptacle, pseudostipe and rhizomorph. For each specimen, 30 basidiospores and 20 hyphae (volva, receptacle, pseudostipe and rhizomorph) were randomly selected and measured. The statistical data used in basidiospores were performed according to Sousa et al. (2014), which determined the variation of the length (H_{Min} and H_{Max}) and width (W_{Min} and W_{Max}), the mean of the two dimensions (x_H and x_W), the standard deviation of these ($\pm SD$) and the height (h) by the width (w) quotient represented by $Q_m [H_{Min}-H_{Max} \times W_{Min}-W_{Max} (x = x_H \pm SD_H \times x_W \pm SD_W, Q_m)]$, always observed under 100 \times objective. The color was coded following Küppers (2002).

Results & Discussion

Taxonomy

Clathrus columnatus Bosc, Mag. Gesell. Naturf. Freunde, Berlin 5: 85 (1811)

Fig. 2a–c

Facesoffungi number: FoF 05677

Description – Immature basidiomata 10–30 mm diam., globose to subglobose, epigeous, white ($N_{00}M_{00}C_{00}$). Expanded basidiomata 30–60 mm high \times 10–25 mm diam. Receptacle orange ($N_{00}Y_{80}M_{50}$), formed by 4 columnar arms, spongy, free at the base, merged at the apex. Volva 10–30 diam., white ($N_{00}Y_{00}M_{00}$), subglobose, glabrous, papery. Gleba covering the inner upper arch of the columns, olive green ($N_{99}C_{80}Y_{80}$), mucilaginous, fetid. Basidiospores 3.3–5.5(–6) \times 1.2–2.5 μ m ($x = 4.5 \pm 0.4 \times 1.9 \pm 0.2$, $Q_m = 2.37$), elongate to cylindrical, hyaline in 5% KOH, smooth. Receptacle composed of pseudoparenchymatous hyphae, 21.5–60 \times 14–42 μ m, globose, subglobose to ovoid, hyaline, thin walls ($<1 \mu$ m). Volva composed of two layers: external layer composed of filamentous hyphae, 2.6–5.4 μ m, straight, with inflated parts, irregular, hyaline, thin walls ($<1 \mu$ m), with septa and branches; gelatinous internal layer, composed of filamentous hyphae, 2.9–4.5 μ m, straight, hyaline, thin walls ($<1 \mu$ m). Rhizomorph composed of filamentous hyphae, 1.3–3.4 μ m, hyaline, thin walls ($<1 \mu$ m), with inflated tips connected by pores.

Habitat – This species has a solitary habit and can be found in soil covered with leaves, sandy soil or decaying wood, in partially or totally shaded areas.

Known distribution – America, Africa and Oceania (Dring 1980). Brazil – Rio de Janeiro (Dring 1980), São Paulo (Bononi et al. 1981), Rio Grande do Sul (Guerrero & Homrich 1999, Sobestiansky 2005, Trierveiler-Pereira et al. 2018), Paraná (Meijer 2006), Paraíba (Magnago et al. 2013) and Ceará (Baseia et al. 2014, present study).

Material examined – Brazil, Ceará, Crato, Araripe National Forest (7°14'55.7"S, 39°29'42.5"W), on soil covered with leaves and decaying wood, 16 Mar 2014, M.M.B. Barbosa, R.H.S.F. Cruz, A100 (UFRN-Fungos 2419). Brazil, Ceará, Crato, Araripe National Forest (7°14'55.7"S, 39°29'42.5"W), on sandy soil, 16 Mar 2014, M.M.B. Barbosa, R.H.S.F. Cruz, A105 (UFRN-Fungos 2424). Brazil, Ceará, Crato, Araripe National Forest (7°14'55.7"S, 39°29'42.5"W), on soil covered with leaves and decaying wood, 17 Mar 2014, M.M.B. Barbosa, R.H.S.F. Cruz, R.J. Ferreira, A115 (UFRN-Fungos 2434).

Notes – Occurring in several parts of Brazil and of the world, this species is common in the Araripe National Forest, Ceará State (Baseia et al. 2014). *Clathrus columnatus* is characterized by presenting basidiomata with 3–4 columnar arms, free at the base and joined at the apex, and gleba scattered at the upper inner part of the arms. It resembles the *Laternea triscapa* Turpin, but the latter presents a gleba confined in a glebifer. *Pseudocolus fusiformis* (E. Fisch.) Lloyd also resembles *C. columnatus*, but the former is distinguished by having arms joined at the base. According to Dring (1980), Cunningham (1944), this species may present regional variants regarding the number of arms, color and shape of the receptacle. In the literature it is recorded

under different names: *Colonnaria* Rafinesque-Schmaltz, *Laternea* Turpin, *Linderia* G. Cunningham, and *Linderiella* G. Cunningham. This is the second record for the Brazilian semi-arid region.

Laternea dringii A. López, D. Martínez & J. García, Boln. Soc. mex. Micol. 16: 110 (1981)

Fig. 2d–f

Facesoffungi number: FoF 05678

Description – Immature basidiomata 8–12 mm high × 7–10 mm diam., subglobose, epigeous, white (N₀₀Y₀₀M₀₀) at the base to yellowish gray (N₆₀C₀₀M₃₀) at the apex, surface glabrous to cracked in the apical portion, not encrusted. Expanded basidiomata 26–40 mm high. Receptacle yellowish white (N₀₀Y₁₀M₀₀) at the base, orange (N₀₀Y₈₀M₅₀) at the apex, formed by 3–4 columnar arms, spongy, free at the base and joined by a transverse arm at the apex, with reniform tubes in transverse section. Absence of transverse arm in some basidiomata. Volva 10–13 mm high × 11–13 wide, white (N₀₀Y₀₀M₀₀) to yellowish gray (N₆₀C₀₀M₃₀), globose to subglobose, surface glabrous to cracked in the apical portion, papery. Rhizomorph up to 40 mm, white (N₀₀Y₀₀M₀₀), adhered at the base of the volva. Glebifer reddish orange (N₀₀Y₉₀M₅₀) in the lower inner part of the arms union. Gleba confined in glebifer, olive green (N₉₀C₁₀Y₉₉), mucilaginous, fetid. Basidiospores 3.5–4.6 × 1.1–2 μm ($x = 4.1 \pm 0.3 \times 1.5 \pm 0.3$, $Q_m = 2.69$), cylindrical to bacilliform, hyaline in 5% KOH, smooth. Receptacle composed of pseudoparenchymatous hyphae, 15.7–48.8 × 14–38.7 μm, subglobose to ellipsoid, hyaline, thin walls (<1 μm). Volva composed of two layers: external layer composed of filamentous hyphae, 3–9 μm, straight, with inflated parts, hyaline, thin walls (<1 μm), with septa and branches; gelatinous internal layer, composed of filamentous hyphae, 2–3.3 μm, straight, hyaline, thin walls (<1 μm), with septa and branches. Rhizomorph composed of filamentous hyphae, 1.8–6.3 μm, hyaline, thin walls (<1 μm), septate, with inflated tips connected by pores.

Habitat – This species has a solitary or gregarious habit and can be found in soil covered with litter in totally shaded area.

Known distribution – Mexico (López et al. 1981, López & García 2001). Brazil – Paraíba (Magnago et al. 2013) and Ceará (present study).

Material examined – Brazil, Ceará, Crato, Alto da Bananeira pub on road (7°14'58.0"S, 39°27'29.9"W), on soil covered with litter, 04 Mar 2018, A.A. Lima, R.A.F. Gurgel, R.L. Oliveira, R.J. Ferreira, CR18-14 (UFRN-Fungos 2985).

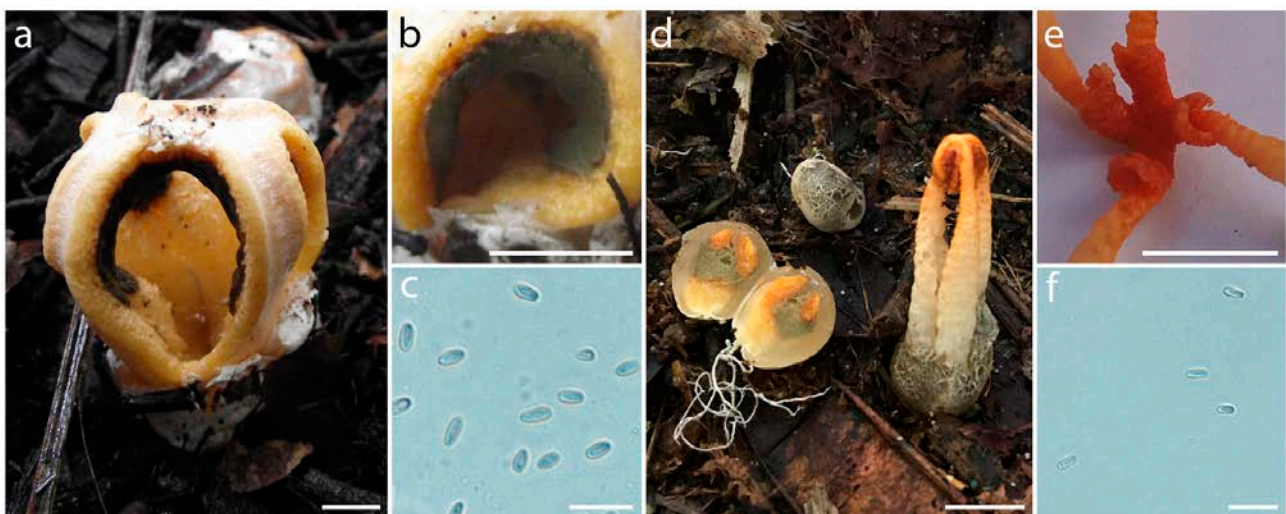


Fig. 2 – *Clathrus columnatus*. a Expanded basidiomata *in situ*. b Gleba covering the inner upper arch of the columns. c Basidiospores. *Laternea dringii*. d Immature and expanded basidiomata *in situ*. e Glebifer in the lower inner part of the arm joints. f Basidiospores. Scale Bars: a, b, d, e = 10 mm; c, f = 10 μm.

Notes – According to López et al. (1981), *Laternea dringii* is characterized by basidiomata 13 to 15 mm in height, red-orange to yellowish orange, formed by 3–4 columns free at the base, joined at the apex by a transverse arm, a glebifer well defined in the interior of the columns and basidiospores measuring 4.2–5(–5.6) x 1.4 µm. The morphological characteristics of the specimens of the present study are very similar to those cited by López et al. (1981). However, present variations: basidiomata larger (26–40 mm high), basidiospores slightly smaller (3.5–4.6 × 1.1–2 µm) and absence of a transverse arm in some basidiomata. It is similar to *Laternea triscapa*, but the latter presents larger basidiomata (up to 70 mm), receptacle vermilion at the apex to pinkish at the base and arms with tubes trapezoid in transverse section (Dring 1980). *Laternea pusilla* Berk. & M.A. Curtis differs from *L. dringii* by presenting columns with crests and receptacle ochraceous to salmon pink. Calonge et al. (2004) considered *L. dringii* a synonym of *L. triscapa*. The present study considered the classification proposed by López et al. (1981). Therefore, more detailed research is needed considering the Brazilian collection, to better delimit the informative characters of this species. In Brazil, Magnago et al. (2013) reported *L. dringii* for Paraíba State. However, only the macroscopic analysis was performed based on a sample that was lost. This is the second record outside Mexico and the first record for the Brazilian semi-arid region.

Mutinus albotruncatus B.D.B. Silva & Baseia, in Silva, Cabral, Martín, Marinho, Calonge & Baseia, Phytotaxa 236: 241 (2015) Fig. 3a–d

Facesoffungi number: FoF 05675

Description – Immature basidiomata 15–18 mm high × 4–7 diam., fabiform to reniform, epigeous, yellowish white (N₀₀Y₁₀M₀₀), with rhizomorph adhered at the base. Expanded basidiomata formed by pseudostipe and volva. Pseudostipe 40–65 mm high × 5–10 mm diam., cylindrical, apically perforate, hollow, spongy; sterile portion white (N₀₀Y₀₀M₀₀), straight, not chambered; fertile portion light brown (N₃₀Y₇₀M₄₀), well-defined, cylindrical, truncate, slightly rugulose, 15–23 mm high, covering about 1/3 of the total length of pseudostipe. Volva 8–20 mm high × 8–10 mm wide, yellowish white (N₀₀Y₁₀M₀₀), with rhizomorph adhered to the base. Rhizomorph up to 40 mm long, yellowish white (N₀₀Y₁₀M₀₀). Gleba covering and confined to the upper region of the pseudostipe, mucilaginous, olive gray (N₉₀C₀₀Y₄₀). Basidiospores 2.8–4.7 × 1.6–3 µm (x = 3.9 ± 0.3 × 2.2 ± 0.2, Q_m = 1.79), ellipsoid to elongate, smooth, hyaline in 5% KOH. Pseudostipe composed of pseudoparenchymatous hyphae, 16.5–48.7 × 12–43 µm hyaline, irregular shaped, thin walls (<1 µm). Volva formed by filamentous hyphae, 2–4.5 µm diam., hyaline, septate, branched. Rhizomorphs composed of filamentous hyphae, 0.3–3.4 µm diam., hyaline, septate.

Habitat – This species has a solitary or gregarious habit and can be found in soil covered with litter or decaying wood, in partially or totally shaded areas.

Known distribution – Brazil – Ceará (Silva et al. 2015, present study).

Material examined – Brazil, Ceará, Crato, Araripe National Forest (7°14'55.7"S, 39°29'42.5"W), on decaying wood, 18 Mar 2014, M.M.B. Barbosa, R.H.S.F. Cruz, A122 (UFRN-Fungos 2441). Brazil, Ceará, Crato, Araripe National Forest (7°14'55.7"S, 39°29'42.5"W), on decaying wood, 19 Mar 2014, M.M.B. Barbosa, R.H.S.F. Cruz, R.J. Ferreira, A136 (UFRN-Fungos 2455). Brazil, Ceará, Crato, Araripe National Forest (7°14'55.7"S, 39°29'42.5"W), on decaying wood, 20 Mar 2014, M.M.B. Barbosa, R.H.S.F. Cruz, A143 (UFRN-Fungos 2461). Brazil, Ceará, Crato, Araripe National Forest, Belmonte trail (7°14'55.7"S, 39°29'42.5"W), on soil covered with litter, 06 Mar 2018, A.A. Lima, R.A.F. Gurgel, R.L. Oliveira, R.J. Ferreira, CR18-39 (UFRN-Fungos 2986).

Notes – This species is common in the Araripe National Forest, Ceará State. This species is characterized by presence of a white pseudostipe, a brown, doliform to cylindrical and truncate fertile portion, and the lignicolous habitat (Silva et al. 2015). Despite having a lignicolous habitat, in our collections it was verified that this species also occurs in soil covered with litter. This species resembles *M. zenkeri* Henn. E. Fischer, but the latter is differentiated by having an incomplete and brittle membrane on fertile portion (Dring & Rose 1977, Degreef et al. 2013). *Mutinus boninensis* Lloyd and *M. borneensis* Lloyd also have a pseudostipe with a white sterile portion. However,

Mutinus boninensis has a fertile portion with a pointed apex, and it is completely annulate (Lloyd 1909, Kobayasi 1937), while *M. borneensis* has a shorter fertile portion, and is salmon pink or red (Lloyd 1909, Grgurinovic 1997). This is the second record for science and the Brazilian semi-arid region.

Mutinus argentinus Speg., Anal. Soc. cient. argent. 24(1): 62 (1887)

Fig. 3e–h

Facesoffungi number: FoF 05676

Description – Immature basidiomata 25 mm high × 13 diam., ellipsoid, semi-hypogeous, white (N₀₀Y₀₀M₀₀) to yellowish brown (N₆₀Y₉₀M₆₀), with rhizomorph adhered at the base. Expanded basidiomata formed by pseudostipe and volva. Pseudostipe 104 mm high × 8 mm diam., cylindrical, apically not perforate, hollow, spongy; sterile portion whitish (N₀₀Y₀₀M₀₀) to light pink (Y₃₀M₄₀C₀₀), slightly curved, not chambered; fertile portion reddish pink (Y₆₀M₉₉C₄₀), well-defined, subulate, slightly rugulose, 30–36 mm high, covering about 1/3 of the total length of pseudostipe. Volva 24 mm high × 14 mm wide, white (N₀₀Y₀₀M₀₀) to yellowish brown (N₆₀Y₉₀M₆₀), with rhizomorph adhered to the base. Rhizomorph up to 70 mm long, white (N₀₀Y₀₀M₀₀). Gleba covering and confined to the upper region of the pseudostipe, mucilaginous, olive brown (N₉₀C₀₀Y₄₀). Basidiospores 3.7–5 × 1–2 μm (x = 4.4 ± 0.3 × 1.4 ± 0.2, Q_m = 3.12), cylindrical to bacilliform, smooth, hyaline in 5% KOH. Pseudostipe composed of pseudoparenchymatous hyphae, 21.8–71 × 13.5–64.5 μm hyaline, irregular shaped, thick walls (>1 μm). Volva formed by filamentous hyphae, 2–3.7 μm diam., hyaline, septate, branched. Rhizomorphs composed of filamentous hyphae, 1.5–7.5 μm diam., hyaline, septate.

Habitat – This species has a gregarious habit and can be found in soil covered with litter in totally shaded areas.

Known distribution – Pantropical (Gube & Piepenbring 2009). Brazil – Paraná (Meijer 2006, Alves & Cortez 2016), Paraíba (Magnago et al. 2013) and Ceará (present study).

Material examined – Brazil, Ceará, Crato, Alto da Bananeira pub on road (7°14'58.0"S, 39°27'29.9"W), on soil covered with litter, 04 Mar 2018, A.A. Lima, R.A.F. Gurgel, R.L. Oliveira, R.J. Ferreira, CR18-13 (UFRN-Fungos 2984).

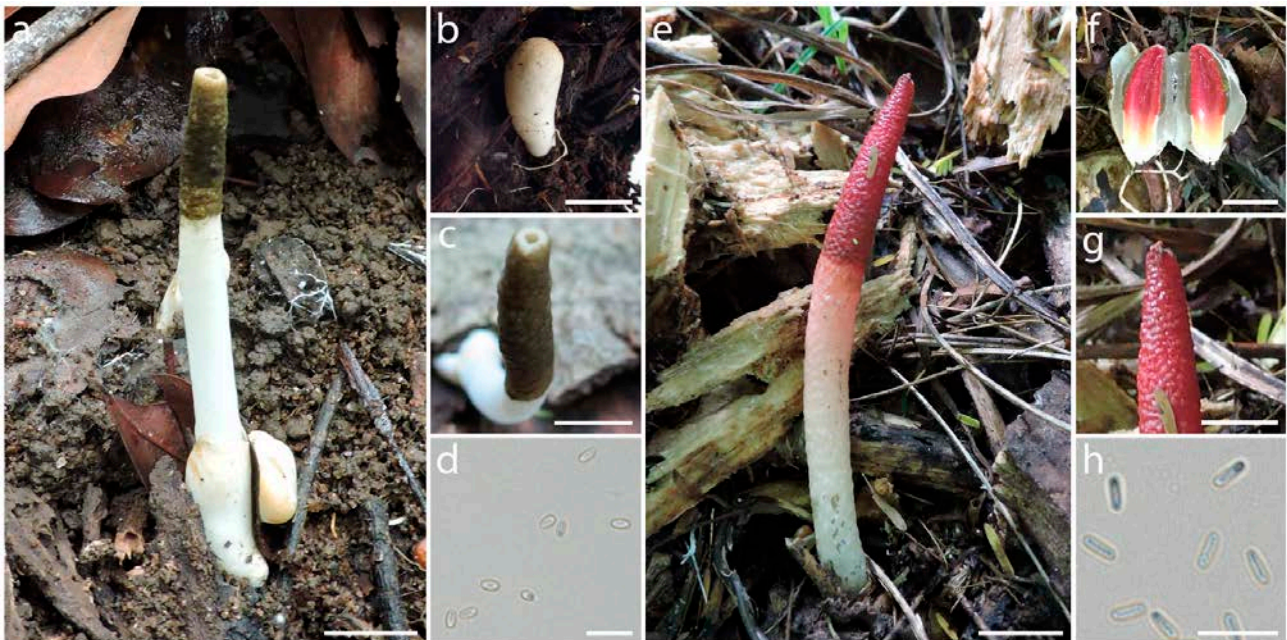


Fig. 3 – *Mutinus albotruncatus*. a Expanded basidioma *in situ*. b Immature basidioma *in situ*. c Fertile portion truncate with perforate apex. d Basidiospores. *Mutinus argentinus*. e Expanded basidioma *in situ*. f Immature basidioma *in situ*. g Fertile portion subulate without perforate apex. h Basidiospores. Scale Bars: a, e, f, g = 10 mm; b, c = 5 mm; d, h = 10 μm.

Notes – This species is recognized by its receptacle with a reddish-pink and well-defined fertile portion, with rugulose surface and pseudostipe the white to light pink color. *Mutinus argentinus* resembles *M. bambusinus* (Zoll.) E. Fisch. However, *Mutinus bambusinus* differs by having a sterile portion occupying 2/3 of the basidiomata, presence of pseudoparenchymatous processes, and a small sterile apical portion with a verrucose surface (Dring & Rose 1977). Due to strong similarities between these taxa, there are some doubts about their taxonomy (Alves & Cortez 2016). Due to their similarity, these two species were treated as synonyms by different authors (Kobayasi et al. 1938, Cunningham 1944, Liu 1984). Dring & Rose (1977) discussed the morphological difference between the two taxa. Gube & Piepenbring (2009) discussed the possibility that some records of *M. bambusinus* in the Neotropics belong to *M. argentinus*. According to Silva et al. (2015), several species of the genus *Mutinus* need to be reviewed. This is the first record for the Brazilian semi-arid region.

Phallus indusiatus Vent., Mém. Inst. nat. Sci. Arts 1: 520 (1798)

Fig. 4a–d

Facesoffungi number: FoF 01080

Description – Immature basidiomata 10–30 mm diam., globose to subglobose, white (N₀₀M₀₀C₀₀) to brown (Y₆₀M₆₀C₅₀). Expanded basidiomata 100–160 mm high. Receptacle 24 mm high × 24 mm wide, white (N₀₀M₀₀C₀₀), campanulate, reticulate surface with apical pore. Pseudostipe 135 mm high × 13 mm wide, white (N₀₀M₀₀C₀₀), cylindrical, spongy, hollow. Indusium well developed extending to the ground, white (N₀₀M₀₀C₀₀), polygonal meshes, joined to the apex of the pseudostipe. Volva 23–30 mm high × 20–35 wide, white (N₀₀M₀₀C₀₀) to brown (Y₆₀M₆₀C₅₀), with pink pigments (N₀₀Y₂₀M₂₀), globose, smooth surface, papery. Rhizomorph white (N₀₀M₀₀C₀₀), adhered to the base of volva. Gleba olive green (N₈₀C₀₀Y₉₉), mucilaginous, fetid. Basidiospores 2.8–4.2 × 1.1–1.9 μm ($x = 3.4 \pm 0.3 \times 1.4 \pm 0.2$, $Q_m = 2.41$), elongated to cylindrical, hyaline in 5% KOH, smooth, thin walls (<1 μm). Pseudostipe composed of pseudoparenchymatous hyphae, 16–40.8 × 14.5–36 μm, globose to subglobose, hyaline, thin walls (<1 μm). Indusium composed of pseudoparenchymatous hyphae, 18.3–19.1 × 19–32 μm, globose to subglobose, hyaline, thin walls (<1 μm). Volva composed of three layers: outer layer composed of filamentous hyphae, 2.5–9.2 μm, straight, with inflated parts, hyaline, thin walls (<1 μm), septate, with connection clamps; intermediate layer gelatinous, composed of filamentous hyphae, 2.1–4.2 μm, straight, hyaline, septate; internal layer composed of filamentous hyphae, 2.5–8 μm, straight, with inflated parts, hyaline, thin walls (<1 μm), septate. Rhizomorph composed of filamentous hyphae, 2.6–9.6 μm, hyaline, thin walls (<1 μm), with inflated tips connected by pores.

Habitat – This species has a solitary habit and can be found in clay or sandy soil covered with litter, in partially shaded areas.

Known distribution – Pantropical (Calonge et al. 2005). Brazil (Möller 1985, Lloyd 1907, Avena-Saccá 1923) – São Paulo (Sydow & Sydow 1907, Viégas 1945, Bononi et al. 1981, Bononi 1984), Rio Grande do Sul (Rick 1961), Paraná (Meijer 2006), Rio Grande do Norte (Baseia et al. 2006), Rondônia (Trierveiler-Pereira et al. 2009), Paraíba (Trierveiler-Pereira & Baseia 2011, Magnago et al. 2013), Espírito Santo, Rio de Janeiro and Santa Catarina (Magnago et al. 2013), Amazonas (Cabral et al. 2014), Mato Grosso do Sul (Bononi et al. 2017) and Ceará (present study).

Material examined – Brazil, Ceará, Crato, Araripe National Forest (7°14'55.7"S, 39°29'42.5"W), on clay soil covered with litter, 20 Mar 2014, M.M.B. Barbosa, R.H.S.F. Cruz, A139 (UFRN-Fungos 2457). Brazil, Ceará, Crato, Araripe National Forest, Belmonte trail (7°14'54.4"S, 39°29'44.2"W), on sandy soil, 06 Mar 2018, A.A. Lima, R.A.F. Gurgel, R.L. Oliveira, R.J. Ferreira, CR18-47 (UFRN-Fungos 2983).

Notes – This species is common in the neotropics (Baseia et al. 2006). *Phallus indusiatus* resembles *P. impudicus* L., distinguished by the absence of indusium in the latter. Other species present indusium as: *Phallus atrovolvatus* Kreisel & Calonge, *P. cinnabarinus* (W.S. Lee) Kreisel, *P. duplicatus* Bosc, *P. echinvolvatus* (M. Zang, D.R. Zheng & Z.X. Hu) Kreisel, *P. luteus* (Liou & L. Hwang) T. Kasuya, *P. merulinus* (Berk.) Cooke, *P. multicolor* (Berk. & Broome) Cooke and *P. rubrovolvatus* (M. Zang, D.G. Ji & X.X. Liu) Kreisel. However, *Phallus indusiatus* is different

from the others because it presents reticulate receptacle, well-developed white indusium, white pseudostipe and volva. *Phallus aureolatus* Trierweiler-Pereira & De Meijer also exhibits white indusium and pseudostipe but differs in its pinkish volva, meruloid to rugulose receptacle and white pseudoparenchymatous cap on the top of the receptacle. Trierweiler-Pereira et al. (2017), in their analyses, indicated that this morphological species probably corresponds to a complex of phylogenetic taxa. *Phallus indusiatus* has been well reported in Brazil. This is the first record for the Brazilian semi-arid region.

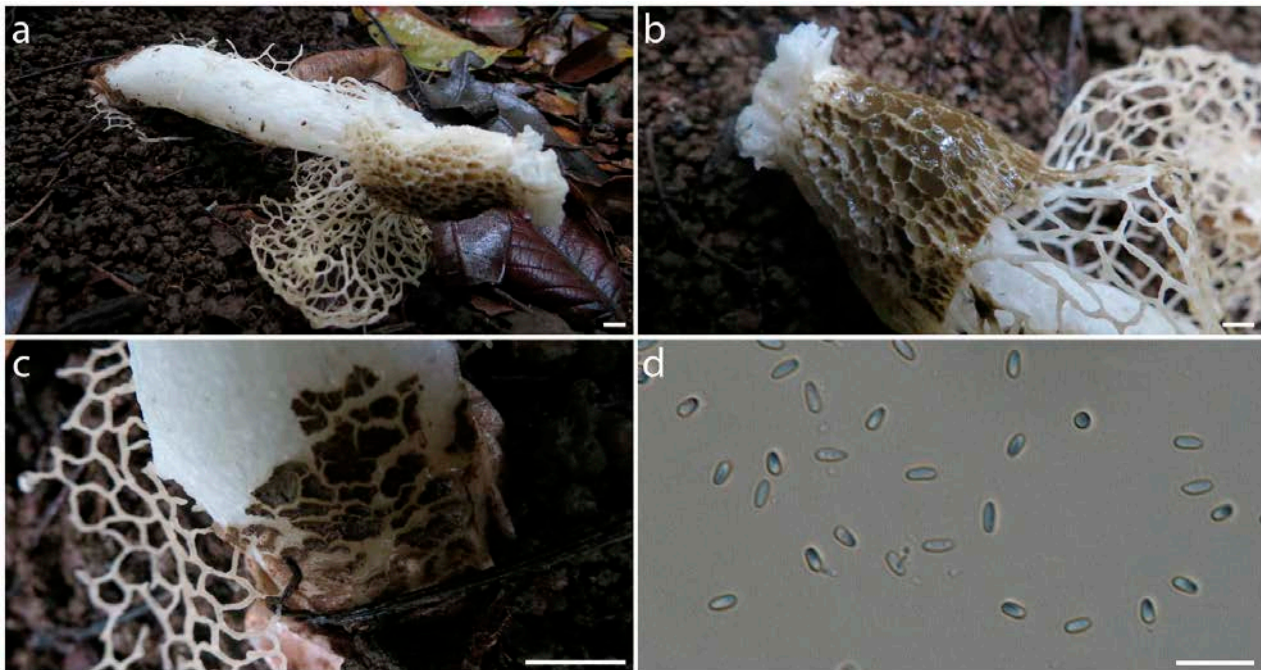


Fig. 4 – *Phallus indusiatus*. a Expanded basidioma *in situ*. b Reticulate receptacle. c Volva. d Basidiospores. Scale Bars: a–c = 10 mm; d = 10 μ m.

Acknowledgements

The authors would like to express their thanks to the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq–Brazil) for the scholarships awarded to Renato J. Ferreira and Rafaela A. F. Gurgel and to the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES–Brazil) for the scholarship awarded to Alexandro A. Lima.

References

- Alves CR, Cortez VG. 2016 – Gasteroid Phallomycetidae (Basidiomycota) from the Parque Estadual São Camilo, Paraná, Brazil. *Iheringia, Série Botânica* 71, 27–42.
- Averna-Saccá R. 1923 – Os gasteromycetes mais comuns nas hortas, nos pomares e nos campos. *Boletim de Agricultura (São Paulo)* 23(9–10), 306–318.
- Baseia IG, Calonge FD, Maia LC. 2006 – Notes on the Phallales in Neotropics. *Boletín de la Sociedad Micológica de Madrid, Spain* 30, 87–93.
- Baseia IG, Silva BDB, Cruz RHSF. 2014 – Fungos Gasteroides no Semiárido do Nordeste Brasileiro. Rio Grande do Norte–Brasil: Print Mídia, 132 pp.
- Bononi VLR. 1984 – Basidiomicetos da Reserva Biológica de Mogi Guaçu. *Rickia* 11, 1–25.
- Bononi VLR, Oliveira AKMD, Gugliotta ADM, Quevedo JRD. 2017 – Agaricomycetes (Basidiomycota, Fungi) diversity in a protected area in the Maracajú Mountains, in the Brazilian central region. *Hoehnea* 44(3), 361–377.
- Bononi VLR, Trufen SFB, Grandi RAP. 1981 – Fungos macroscópicos do Parque Estadual das Fontes do Ipiranga depositados no Herbário do instituto de Botânica. *Rickia* 9, 37–53.

- Cabral TS, Silva BDB, Ishikawa NK, Alfredo DS et al. 2014 – A new species and new records of gasteroid fungi (Basidiomycota) from Central Amazonia, Brazil. *Phytotaxa* 183(4), 239–253.
- Calonge FD, Guzmán G, Ramírez-Guillén F. 2004 – Observaciones sobre los Gasteromycetes de México depositados en los herbários XAL y XALU. *Boletín de la Sociedad Micológica de Madrid* 28, 337–371.
- Calonge FD, Mata M, Carranza J. 2005 – Contribución al catálogo de los gasteromycetes (basidiomycotina, fungi) de Costa Rica. *Anales del Jardín Botánico de Madrid* 62, 23–45.
- Cortez V, Baseia IG, Silveira RMB. 2011a – Two noteworthy *Phallus* from southern Brazil. *Mycoscience* 52, 436–438.
- Cortez V, Baseia IG, Silveira RMB. 2011b – Gasteroid mycobiota of Rio Grande do Sul, Brazil: Lysuraceae (basidiomycota). *Acta Scientiarum Biological Sciences* 33, 87–92.
- Crous PW, Wingfield MJ, Burgess TI, Carnegie AJ et al. 2017 – Fungal Planet description sheets: 625–715. *Persoonia: Molecular Phylogeny and Evolution of Fungi* 39, 270–467.
- Cunningham CH. 1944 – The Gasteromycetes of Australia and New Zealand. Dunedin, 236 pp.
- Degreef J, Amalfi M, Decock C, Demoulin V. 2013 – Two rare Phallales recorded from São Tomé. *Cryptogamie, Mycologie* 34 (1), 3–13.
- Dring DM. 1980 – Contributions towards a rational arrangement of the Clathraceae. *Kew Bulletin [S.l.]* 35(1), 1–96.
- Dring DM, Rose AC. 1977 – Additions to West African phalloid fungi. *Kew Bulletin* 31, 741–751.
- Fazolino EP, Trierveiler-Pereira L, Calonge FD, Baseia IG. 2010 – First records of *Clathrus* (Phallaceae, Agaricomycetes) from the northeast region of Brazil. *Mycotaxon* 113, 195–202.
- Grgurinovic AC. 1997 – Larger Fungi of South Australia. Botanic Gardens of Adelaide and State Herbarium and Flora and Fauna of South Australia Handbooks Committee, Adelaide, 765 pp.
- Gube M, Piepenbring M. 2009 – Preliminary annotated checklist of gasteromycetes in Panama. *Nova Hedwigia* 89, 519–543.
- Guerrero RT, Homrich MH. 1999 – Fungos Macroscópicos Comuns no Rio Grande do Sul. Guia para identificação. Ed. Universidade/UFRGS: Porto Alegre (Brasil). 124 pp.
- Hibbett DS, Bauer R, Binder M, Giachini AJ et al. 2014 – 14 Agaricomycetes. *Systematics and Evolution: The Mycota* 7(2), 373–429.
- Hosaka K, Bates ST, Beaver ER, Castellano MA et al. 2006 – Molecular phylogenetics of the gomphoid-phalloid fungi with an establishment of the new subclass Phallomycetidae and two new orders. *Mycologia* 98(6), 949–959.
- IBGE. 2010 – Instituto Brasileiro de Geografia e Estatística. Censo demográfico 2010. <https://www.ibge.gov.br/cidadesat> (accessed 26 Apr 2018).
- Kobayasi Y. 1937 – Fungi Austro-Japoniae et Micronesiae. I. *The Botanical Magazine* 51, 749–758.
- Kobayasi Y. 1938 – Hymenogastrineae et phallineae. In T. Nakai and M. Honda [eds.], *Nova Flora Japonica*. Tokyo and Osaka. Sanseido, 1–90.
- Küppers H. 2002 – Atlas de los colores. 1 st ed. Barcelona: Blume, 165 pp.
- Leal IR, Silva JMC, Tabarelli M, Lacher Jr TE. 2005 – Mudando o curso da conservação da biodiversidade na Caatinga do Nordeste do Brasil. *Megadiversidade* 1, 139–146.
- Liu B. 1984 – The gasteromycetes of China. *Beihefte zur Nova Hedwigia* 74, 1–235.
- Lloyd CG. 1907 – Mycological Notes n° 26. Concerning the phalloids. *Mycology Writings* 2, 325–337.
- Lloyd CG. 1909 – Mycological Notes n° 4. Synopsis of the Know Phalloids. *Mycology Writings* 4, 1–96.
- López A, García J. 2001 – *Laternea dringii*. Instituto de Genética Forestal, Universidad Veracruzana, Xalapa, Vera Cruz, México. *Funga Veracruzana* 36, 1–2.
- López A, Martínez D, García J. 1981 – Adiciones al conocimiento de los Phallales del estado de Veracruz. *Boletín de la Sociedad Mexicana de Micología* 16, 109–116.

- Magnago AC, Trierveiler-Pereira L, Neves MA. 2013 – Phallales (Agaricomycetes, Fungi) from the tropical Atlantic Forest of Brazil. *Journal of the Torrey Botanical Society* 140 (2), 236–244.
- Meijer AAR. 2006 – Preliminary list of the macromycetes from the Brazilian State of Paraná. *Boletim do Museu Botânico Municipal* 68, 1–55.
- MMA. 2006 – Ministério do Meio Ambiente. Plano operativo de prevenção e combate aos incêndios florestais da Floresta Nacional de Araripe-Apodi. 2006. <http://www.ibama.gov.br/phocadownload/category/44-p?download=2325%3ap> (accessed 5 May 2018).
- Möller A. 1895 – Brasilische Pilzblumen. *Botanische Mittheilungen aus den Tropen* 7, 1–152.
- Pereira-junior JS. 2007 – Nova delimitação do Semiárido brasileiro. Brasília, DF, 32 pp.
- Rick J. 1961 – Basidiomycetes Eubasidii no Rio Grande do Sul. Brasília. *Iheringia* 9, 451–480.
- Silva BDB, Cabral TS, Martín MP, Marinho P et al. 2015 – *Mutinus albo truncatus* (Phallales, Agaricomycetes), a new phalloid from the Brazilian semiarid, and a key to the world species. *Phytotaxa* 236(3), 237–248.
- Sobestiansky G. 2005 – Contribution to a Macromycete survey of the states of Rio Grande do Sul and Santa Catarina in Brazil. *Brazilian Archives of Biology and Technology* 48, 437–457.
- Sousa JO, Silva BDB, Baseia IG. 2014 – *Geastrum* from the Atlantic Forest in northeast Brazil — new records for Brazil. *Mycotaxon* 129, 169–179.
- Sulzbacher MA, Grenbec T, Cabral TS, Giachini AJ et al. 2016 – *Restingomyces*, a new sequestrate genus from the Brazilian Atlantic rainforest that is phylogenetically related to early-diverging taxa in Trappeaceae (Phallales). *Mycologia* 108(5), 954–966.
- Sydow H, Sydow P. 1907 – Verzeichnis der von Herrn F. Noack in Brasilien gesammelten Pilze. *Annales Mycologici* 5(4), 348–363.
- Trierveiler-Pereira L, Baseia IG. 2011 – Contribution to the knowledge of gasteroid fungi (Agaricomycetes, Basidiomycota) from the state of Paraíba, Brazil. *Revista Brasileira de Biociências* 9(2), 167–173.
- Trierveiler-Pereira L, Gomes-Silva AC, Baseia IG. 2009 – Notes on gasteroid fungi of the Brazilian Amazon rainforest. *Mycotaxon* 110, 73–80.
- Trierveiler-Pereira L, Honaiser LP, da Silveira RMB. 2018 – Diversity of gasteroid fungi (Agaricomycetes, Basidiomycota) from the Brazilian Pampa Biome. *Nova Hedwigia* 105(3–4), 3–4.
- Trierveiler-Pereira L, Meijer AAR, Reck MA, Hosaka K, Silveira RMB. 2017 – *Phallus aureolatus* (Phallaceae, Agaricomycetes), a new species from the Brazilian Atlantic Forest. *Phytotaxa* 327(3), 223–236.
- Tuno N. 1998 – Spore dispersal of *Dictyophora* fungi (Phallaceae) by flies. *Ecological Research* 13, 7–15.
- Viégas AP. 1945 – Alguns fungos do Brasil, 10: Gasteromicetos. *Bragantia*, 5(9), 583–595.